

VERIFICATION OF TRANSLATION

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declare as follows:

1. That I am well acquainted with both the English and Japanese languages, and
2. That the attached document is a true and correct translation of a certified copy of the following application, which was made by me to the best of my knowledge and belief:
 - (a) Japanese Patent Application No. 2002-339418
Entitled: "METHOD OF SCREENING FOR COMPOUNDS THAT INHIBIT THE
ENZYMATIC ACTIVITY OF GWT1 GENE PRODUCT"
Filed on November 22, 2002

April 25, 2006
(Date)

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【Name of Document】 APPLICATION FOR PATENT

【Identification Number】 E1-A0209

【Filing Date】 November 22, 2002

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【Payment】

【Registration Number】 041092

【Amount】 10,500

【List of Attached Documents】

【Name of Document】 Description 1

【Name of Document】 Drawings 1

【Name of Document】 Abstract 1

【Proof】 Necessary

[Document Name] Specification

[Title of the Invention] METHOD OF SCREENING FOR COMPOUNDS THAT INHIBIT THE ENZYMATIC ACTIVITY OF GWT1 GENE PRODUCT

5

[Claims]

[Claim 1] A method of screening for a compound having an antifungal activity, wherein the method comprises the steps of:

- 10 (1) contacting a test sample with an overexpressed protein encoded by the GWT1 gene;
(2) detecting GlcN-(acyl)PI; and
(3) selecting the test sample that decreases GlcN-(acyl)PI.

[Claim 2] The method of claim 1, wherein the GWT1 gene is any one of the following:

- 15 (a) a DNA encoding a protein comprising the amino acid sequence of SEQ ID NO: 2, 4, 6, 8, 10, or 14;
(b) a DNA comprising the nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 9, 11, 12, or 13;
(c) a DNA hybridizing to the DNA comprising the nucleotide sequence of SEQ ID NO:
20 1, 3, 5, 7, 9, 11, 12, or 13 under stringent conditions; and
(d) a DNA encoding a protein comprising the amino acid sequence of SEQ ID NO: 2, 4, 6, 8, 10, or 14, wherein one or more amino acids have been added, deleted, substituted, and/or inserted.

25 [Claim 3] The method of claim 1 or 2, wherein the step of detecting the acylated GPI is thin-layer chromatography.

[Claim 4] The method of any one of claims 1 to 3, wherein the method further comprises a step
4, of determining whether the selected test sample inhibits the process of transporting a
30 GPI-anchored protein to a fungal cell wall, whether the test sample inhibits the expression of a GPI-anchored protein on a fungal cell surface, or whether the test sample inhibits the proliferation of a fungi.

[Detailed Description of the Invention]

35 [0001] [Technical Field of Industrial Application]

The present invention relates to methods of screening for antifungal agents having the

activity of inhibiting GPI synthase, which is involved in the synthesis of fungal cell walls.

[0002] [Prior Art]

The present inventors noticed that adhesion to host cells is important for fungi to exert their pathogenicity, and that adhesion factors involved in fungal cell adhesion are transported to the surface layers of cell walls after glycosylphosphatidylinositol (GPI) anchors on the cell membrane (Non-Patent Document 1). Accordingly, the present inventors considered that novel antifungal agents that inhibit the synthesis of fungal cell walls and also inhibit the adhesion of fungal cells to host cells could be generated by inhibiting the process of transporting proteins anchored with GPI (GPI-anchored proteins) to cell walls. Thus, the present inventors started study.

[0003] The prior art reference related to the invention of the present application is shown below:

[Non-Patent Document 1] Hamada K *et al.*, Mol. Gen. Genet., 258: 53-59, 1998

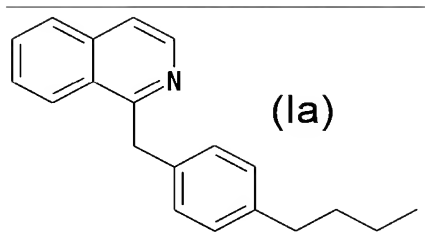
[0004] [Problems to be Solved by the Invention]

An objective of the present invention is to develop antifungal agents for preventing pathogenic fungi from exerting pathogenicity, by inhibiting the synthesis of fungal cell walls, as well as by inhibiting fungal cell adhesion to host cells, by inhibition of the transport of GPI-anchored proteins to fungal cell walls.

[0005] [Means to Solve the Problems]

In WO 02/04626, the present inventors found the following proteins involved in the process of transporting GPI-anchored proteins to cell walls: the proteins of *Saccharomyces cerevisiae* encoded by DNAs comprising the nucleotide sequence of SEQ ID NO: 1; the proteins of *Candida albicans* encoded by DNAs comprising the nucleotide sequences of SEQ ID NOs: 3 and 5; the proteins of *Schizosaccharomyces pombe* encoded by DNAs comprising the nucleotide sequence of SEQ ID NO: 7; the proteins of *Aspergillus fumigatus* encoded by DNAs comprising the nucleotide sequences of SEQ ID NOs: 9 and 11; and the proteins of *Cryptococcus neoformans* encoded by DNAs comprising the nucleotide sequences of SEQ ID NOs: 12 and 13. These nucleotide sequences were called GWT1 genes. In addition, the inventors found that GWT1 gene-deficient fungi can not synthesize cell walls. Furthermore, the inventors found that the compound represented by formula (Ia) binds to the above-described proteins to inhibit the transport of GPI-anchored proteins to cell walls, thus inhibiting the synthesis of fungal cell walls.

[0006] [Compound 1]



[0007] The inventors then found that the GWT1 gene product (hereinafter referred to as “GWT1 protein”) has the activity of synthesizing GlcN-(acyl)PI by transferring an acyl group to GlcN-PI in the GPI biosynthesis pathway (Fig. 1; Kinoshita and Inoue, Curr Opin Chem Biol 2000 Dec;4(6): 632-8; Ferguson *et al.*, Biochim Biophys Acta 1999 Oct 8; 1455 (2-3): 327-40). The inventors conceived that compounds inhibiting the synthesis of fungal cell walls could be found by screening for compounds that inhibit this activity, and thus completed the present invention.

[0008] Specifically, the present invention provides 1 to 7 as described below.

1. A method of screening for a compound having an antifungal activity, wherein the method comprises the steps of:

- (1) contacting a test sample with an overexpressed protein encoded by the GWT1 gene;
- (2) detecting GlcN-(acyl)PI; and
- (3) selecting the test sample that decreases GlcN-(acyl)PI.

[0009] The “GWT1” gene refers to a gene involved in the synthesis of fungal cell walls, which was disclosed in WO 02/04626. The term “overexpressed” does not refer to expression of native genes, but to the expression of exogenously introduced genes.

[0010] “GlcN-(acyl)PI” refers to glucosaminyl-acylphosphatidylinositol in which an acyl group is linked with the inositol of glucosaminyl-phosphatidylinositol (GlcN-PI) in the GPI biosynthesis pathway (Fig. 1; Kinoshita and Inoue, Curr Opin Chem Biol 2000 Dec; 4(6):632-8; Ferguson *et al.*, Biochim Biophys Acta 1999 Oct 8; 1455(2-3):327-40).

[0011] 2. The method of claim 1, wherein the GWT1 gene is any one of the following:

- (a) a DNA encoding a protein comprising the amino acid sequence of SEQ ID NO: 2, 4, 6, 8, 10, or 14;
- (b) a DNA comprising the nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 9, 11, 12, or 13;
- (c) a DNA hybridizing to the DNA comprising the nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 9, 11, 12, or 13 under stringent conditions; and
- (d) a DNA encoding a protein comprising the amino acid sequence of SEQ ID NO: 2, 4, 6, 8, 10, or 14, wherein one or more amino acids have been added, deleted, substituted, and/or inserted.

[0012] The term “stringent conditions” means, for example, hybridization in 4x SSC at 65°C followed by washing with 0.1x SSC at 65°C for one hour. Alternatively, stringent conditions refer to hybridization in 4x SSC with 50% formamide at 42°C. Other acceptable conditions may be hybridization in PerfectHyb™ solution (TOYOBO) at 65°C for 2.5 hours, followed by washing with (1) 2x SSC, 0.05% SDS at 25°C for five minutes; (2) 2x SSC, 0.05% SDS at 25°C for 15 minutes; and (3) 0.1x SSC, 0.1% SDS at 50°C for 20 minutes.

[0013] The “protein comprising an amino acid sequence in which one or more amino acids have been added, deleted, substituted, and/or inserted” can be prepared by methods known to those skilled in the art, for example, by site-directed mutagenesis (Sambrook, J., Fritsch, E. F., and Maniatis, T. (1989) *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY). Such mutations can also occur naturally. There is no limitation on the number of amino acids to be mutated, as long as the binding activity with the labeled compound is maintained. The number of amino acids to be mutated is typically 30 or less, preferably ten or less, and more preferably three or less. There is no limitation on the position of the mutated amino acids, as long as the protein retains the activity described above.

[0014] The proteins and protein mutants prepared using the above-described hybridization techniques normally have high homology (for example, 60% or higher, 70% or higher, 80% or higher, 90% or higher, or 95% or higher homology) to proteins consisting of the amino acid sequence of SEQ ID NO: 2, 4, 6, 8, 10, or 14 at the amino acid level. The amino acid sequence homology can be determined using a BLASTx program (at the amino acid level; Altschul *et al.*, J. Mol. Biol. 215:403-410, 1990). This program is based on the BLAST algorithm of Karlin and Altschul (Proc. Natl. Acad. Sci. USA 87:2264-2268, 1990; Proc. Natl. Acad. Sci. USA 90:5873-5877, 1993). When the amino acid sequences are analyzed using BLASTX, parameters of, for example, score= 50 and wordlength= 3 are used. Alternatively, when using the Gapped BLAST program, the amino acid sequences can be analyzed by the method described by Altschul *et al.* (Nucleic. Acids. Res. 25:3389-3402, 1997). When the BLAST and Gapped BLAST programs are used, the default parameter values for each program are used. Specific procedures for these analyses are known in the art (<http://www.ncbi.nlm.nih.gov>).

[0015] 3. The method of claim 1 or 2, wherein the step of detecting the acylated GPI is thin-layer chromatography.

4. The method of any one of claims 1 to 3, wherein the method further comprises a step 4, of determining whether the selected test sample inhibits the process of transporting a GPI-anchored protein to a fungal cell wall, whether the test sample inhibits the expression of a GPI-anchored protein on a fungal cell surface, or whether the test sample inhibits the proliferation of a fungi.

[[0016] [Mode for Carrying Out the Invention]

Methods for preparing GWT1 protein [1], and methods for determining transacylation activity [2] of the present invention are disclosed below.

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1. Methods for preparing GWT1 protein

GWT1 protein is prepared from a fungal membrane fraction, preferably that of *S. cerevisiae*, *C. albicans*, *S. pombe*, *A. fumigatus*, or *C. neoformans*, and more preferably *S. cerevisiae*. The transacylation activity may be determined by using the prepared membrane fraction directly or after purification. The transacylation activity can be readily measured by introducing a DNA of the nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 9, 11, 12, or 13 into fungal cells to overexpress the GWT1 protein. This procedure can be specifically described using *S. cerevisiae*, as follows:

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[0017] (1) Introduction of the GWT1 gene

The GWT1 gene can be prepared by carrying out PCR using fungal DNAs as templates, and primers designed based on a nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 9, 11, 12, or 13.

20

The GWT1 expression plasmid is prepared by inserting an appropriate promotor and terminator, such as a GAPDH promoter and a GAPDH terminator derived from pKT10 (Tanaka *et al.*, Mol. Cell Biol., 10: 4303-4313, 1990), into the multi-cloning site of an expression vector that functions in *S. cerevisiae*, such as YEp352, and inserting the GWT1 gene into the expression vector. *S. cerevisiae* cells of, for example, G2-10 strain, are incubated while shaking in an appropriate medium such as yeast extract-polypeptone-dextrose (YPD) medium at an appropriate temperature, for example, at 30°C. The fungal cells are harvested at the late logarithmic growth phase. After washing, GWT1 expression plasmids are introduced into *S. cerevisiae* cells, for example, by the lithium acetate method. The lithium acetate method is described in the Users Manual attached to YEAST MAKER™ Yeast Transformation System (Clontech). GWT1-overexpressing strain and empty vector-introduced strain can be obtained by culturing the cells in SD(ura-) medium at 30°C for two days.

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[0018] Fungal strains to which the GWT1 gene is introduced are preferably deficient strains lacking their native GWT1 gene. *S. cerevisiae* GWT1 gene-deficient cells can be obtained by a method described below.

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PCR amplification is carried out using a marker gene, preferably *S. pombe* *his5* gene, as a template, and primers designed to obtain PCR products which comprise 30 bp, or more preferably 40 bp or more of the GWT1 gene sequence (for example, the sequence of SEQ ID NO: 1) to be deleted. The resulting PCR products are purified, and then introduced into fungal

cells. Deficient strains can be obtained by screening appropriate to the marker gene, for example, by culturing the cells in his- medium when the marker is his5.

[0019] Expression vectors and gene introduction methods for fungus other than *S. cerevisiae* are described in: Igarashi *et al.*, Nature 353: 80-83, 1991, for *S. pombe* expression vector pcL and such, and methods for introducing the vectors; Pla J *et al.*, Yeast, 12: 1677-1702, 1996, for *C. albicans* expression vector pRM10 and such, and methods for introducing these vectors; Punt PJ *et al.*, GENE, 56: 117-124, 1987, for *A. fumigatus* expression vector pAN7-1 and such, and methods for introducing these vectors; and Monden P *et al.*, FEMS Microbiol. Lett., 187: 41-45, 2000, for *C. neoformans* expression vector pPM8 and such, and methods for introducing these vectors.

Methods for preparing deficient strains of *C. albicans* are described in Fonzi WA *et al.*, Genetics 134: 717-728, 1993.

[0020] (2) Methods for preparing the membrane fraction

S. cerevisiae cells to which the GWT1 gene are introduced are cultured while shaking in an appropriate medium, such as SD(ura-) liquid medium, at an appropriate temperature, for example 24°C. The fungal cells are harvested in the middle logarithmic growth phase. After being washed with TM buffer (50 mM Tris-HCl (pH 7.5) and 2 mM MgCl₂), the fungal cells are suspended in an adequate amount (for example, 2 ml) of TM buffer + protease inhibitor (CompleteTM; Roche). An adequate amount (for example, 1.5 ml) of glass beads is added to the suspension. The samples are vortexed and placed on ice, and these procedures are repeated (for example, ten cycles of vortexing for 30 seconds and placing on ice for 30 seconds) to disrupt fungal cells.

The samples are centrifuged, for example, at 1000 g for five minutes, to precipitate glass beads and fungal cells which are not disrupted. The resulting supernatant is transferred to another tube, and then centrifuged, to precipitate the membrane fraction comprising organelles (total membrane fraction), for example at 13 000 g for 20 minutes. If required, the precipitate is further suspended in 1 ml of an appropriate assay buffer, and centrifuged, for example, at 1000 g for one minute to remove those components which are not suspended. The supernatant is then centrifuged, for example, at 13 000 g for 20 minutes, and the resulting precipitate is resuspended in an appropriate assay buffer to obtain a membrane fraction.

[0021] Membrane fractions from fungal cells other than *S. cerevisiae* can be prepared by the methods as described in: Yoko-o *et al.*, Eur. J. Biochem. 257: 630-637, 1998, for *S. pombe*; Sentandreu M *et al.*, J. Bacteriol., 180: 282-289, 1998, for *C. albicans*; Mouyna I *et al.*, J. Biol. Chem., 275: 14882-14889, 2000, for *A. fumigatus*; and Thompson JR *et al.*, J. Bacteriol., 181: 444-453, 1999, for *C. neoformans*.

[0022] Alternatively, GWT1 protein can be prepared by expression in cells other than fungal cells, such as mammalian cells, insect cells, and *E. coli* cells.

When mammalian cells are used, a membrane fraction can be prepared by inserting GWT1 into, for example, an overexpression vector comprising CMV promotor; introducing the vector into mammalian cells; and then carrying out the method described in Petaja-Repo *et al.*, J. Biol. Chem., 276: 4416-23, 2001.

[0023] When insect cells are used, a membrane fraction can be prepared by preparing GWT1-expressing insect cells (such as Sf9 cells) using a baculovirus expression kit, for example, BAC-TO-BAC Baculovirus Expression system (GIBCO BRL); and then using the cells to carry out the method described in Okamoto *et al.*, J. Biol. Chem., 276: 742-751, 2001.

When *E. coli* is used, GWT1 protein can be prepared by inserting GWT1 into an *E. coli* expression vector, for example, pGEX (Pharmacia); and then introducing the vector into *E. coli* cells such as BL21.

[0024] 2. Methods for determining transacylation activity

The transacylation reaction to GPI can be detected by the method described in Costello and Orlean, J. Biol. Chem. (1992) 267: 8599-8603, or the method described in Franzot and Doering, Biochem. J. (1999) 340: 25-32. Examples of specific methods are illustrated below, however, the experimental conditions below are preferably optimized according to the GWT1 gene products to be used, as follows:

The GWT1 gene product prepared in Section 1, above, preferably a membrane fraction comprising a GWT1 gene product, is added along with test compounds to a buffer comprising: appropriate metal ions (Mg, Mn); ATP; and Coenzyme A; and preferably inhibitors that prevent the consumption of UDP-GlcNAc in other reactions, such as nikkomycin Z as an inhibitor of chitin synthesis, and tunicamycin as an inhibitor of the synthesis of asparagine-linked sugar chain. The mixture is incubated at an appropriate temperature for an appropriate period (for example, at 24°C for 15 minutes).

[0025] Then, a GlcN-(acyl)PI precursor (for example, UDP-GlcNAc or Acyl-Coenzyme A, and preferably UDP-[¹⁴C]GlcNAc) labeled with an appropriate label, preferably with an isotope, is added to the mixture. The resulting mixture is further incubated for an appropriate period (for example, for one hour at 24°C). A 1:2 mixture of chloroform: methanol is added to the mixture, and stirred to stop the reaction. Lipids are then extracted from the mixture. The extracted reaction products are dissolved in an appropriate solvent, preferably in butanol, and then subjected to HPLC, thin-layer chromatography (TLC), or such, and preferably TLC, to isolate GlcN-(acyl)PI generated in the reaction. A developing solvent for TLC can be selected appropriately, and may be, for example, CHCl₃/CH₃OH/H₂O (65:25:4), CHCl₃/CH₃OH/1 M

NH₄OH (10:10:3), or CHCl₃/pyridine/HCOOH (35:30:7), and preferably HCl₃/CH₃OH/1 M NH₄OH (10:10:3). The isolated GlcN-(acyl)PI is quantified by a method that accords with the label used. When labeled with an isotope, the isolated GlcN-(acyl)PI is quantified based on its radioactivity.

5 When a reduced amount of GlcN-(acyl)PI is produced in the presence of a test compound, the test compound is determined to comprise the activity of inhibiting transacylation by GWT1 proteins.

[0026] A test sample found to comprise the activity of inhibiting transacylation as described above, is preferably further tested to determine whether it inhibits the process of
10 transporting GPI-anchored proteins to fungal cell walls, whether it inhibits the expression of GPI-anchored proteins on fungal cell surfaces, or whether it inhibits fungal growth. If the test results show that the test sample inhibits the process of transporting GPI-anchored proteins to fungal cell walls, inhibits the expression of GPI-anchored proteins on fungal cell surfaces, or inhibits fungal growth, then the sample is a promising candidate for an antifungal agent.

15 [0027] Methods that (1) use reporter enzymes; (2) use antibodies that react to glycoproteins on the surface layers of fungal cell walls; (3) test fungal cells for adhesiveness to animal cells; or (4) observe fungal cells under a light microscope or electron microscope can be used to test whether a test sample inhibits the process of transporting GPI-anchored proteins to fungal cell walls or inhibits the expression of GPI-anchored proteins on fungal cell surfaces.

20 [0028] Methods (1) to (4) are enclosed in WO 02/04626, and specifically illustrated in the Examples. By using the methods of (1) to (4), preferably in combination, a test sample can be determined to inhibit the process of transporting GPI-anchored proteins to fungal cell walls or to inhibit the expression of GPI-anchored proteins on fungal cell surfaces. Further, a test sample can be determined to effect the process of transporting GPI-anchored proteins to cell
25 walls, when the inhibition by the test sample is impaired or disappears when a protein encoded by a DNA of the present invention is overexpressed in fungal cells.

[0029] Conventional methods for measuring antifungal activity can also be used to determine whether a test sample inhibits fungal growth (National Committee for Clinical Laboratory Standards. 1992. Reference method for broth dilution antifungal susceptibility testing
30 for yeasts. Proposed standard M27-P. National Committee for Clinical Laboratory Standards, Villanova, Pa.).

[0030] [Examples]

Herein below, the present invention will be specifically described using Examples, but it
35 is not to be construed as being limited thereto.

[Example 1] Preparation of membrane fraction expressing GWT1 protein

(1) Preparation of GWT1 expression plasmid

The vector for expressing in *S. cerevisiae*, YEp352GAPII vector, was prepared by inserting a GAPDH promoter and a GAPDH terminator, both derived from pKT10 (Tanaka *et al.*,
5 Mol. Cell Biol., 10: 4303-4313, 1990), into the multi-cloning site of YEp352; and replacing the multi-cloning site with that of pUC18. Furthermore, to facilitate the insertion of the GWT1 gene, YEp352GAPIIClaIΔSal vector was prepared by substituting the ClaI site for the SalI site in the multi-cloning site.

10 The *S. cerevisiae* GWT1 gene comprising the nucleotide sequence of SEQ ID NO: 1 was amplified using the primers of SEQ ID NOs: 15 and 16. The resulting PCR product was inserted into the multi-cloning site of YEp352GAPIIClaIΔSal vector to prepare the GWT1 overexpression plasmid.

[0031] (2) Preparation of *S. cerevisiae* GWT1 gene-deficient strain Δgwt1

15 A his5 cassette comprising GWT1 sequences at both ends was amplified by PCR using the *S. pombe* his5 gene (Longtine MS *et al.*, Yeast, 14: 953-961, 1998) as a template and the sequences of SEQ ID NOs: 17 and 18 as primers.

S. cerevisiae cells were cultured and harvested, and then subjected to transformation with the PCR products described above. Then, the cells were cultured in SD(His-) medium at
20 30°C for five to seven days to obtain GWT1 gene-deficient strain Δgwt1.

[0032] (3) Preparation of GWT1-expressing cells

Cells of the Δgwt1 strain were cultured while shaking in yeast extract-polypeptone-dextrose (YPD) medium at 30°C. The cells were harvested in the late
25 logarithmic growth phase and then washed. The expression plasmid for GWT1 was introduced to the Δgwt1 strain cells by the lithium acetate method (YEAST MAKER™ Yeast Transformation System (Clontech)). Δgwt1 strain overexpressing the GWT1 gene was obtained by culturing the cells in SD(ura-) medium at 30°C for two days.

30 [0033] (4) Preparation of membrane fraction

Wild-type *S. cerevisiae* strain, the GWT1 gene-deficient strain Δgwt1, and the strain Δgwt1 into which the GWT1 overexpression plasmid was introduced were each cultured in 100 ml of YPD medium shaken at 24°C, and then harvested in the middle logarithmic growth phase (OD₆₀₀= 1 ~ 3). The fungal cells were washed with TM buffer (50 mM Tris-HCl (pH 7.5) and 2
35 mM MgCl₂), and then suspended in 2 ml of TM buffer + protease inhibitor (1 tablet of Complete™ (Roche) / 25 ml). 1.5 ml of glass beads was added to the suspension. The

mixture was vortexed for 30 seconds, and then placed on ice for 30 seconds. These procedures were repeated ten times to disrupt the fungal cells. The cell homogenate was transferred into a new tube, and centrifuged at 1000 g at 4°C for five minutes to precipitate the glass beads and undisrupted fungal cells. The supernatant was transferred to another tube, and centrifuged at 13000g at 4°C for 20 minutes to precipitate the membrane fraction comprising organelles (total membrane fraction). The resulting precipitate was used as the membrane fraction.

[0034] (5) Detection of acylated GPI

In the GPI biosynthesis reaction pathway, it is known that

N-acetyl-glucosaminyl-phosphatidylinositol (GlcNAc-PI) is deacetylated to generate glucosaminyl-phosphatidylinositol (GlcN-PI), to which an acyl group is then added to generate glucosaminyl-acylphosphatidylinositol (GlcN-(acyl)PI) (Fig. 1). The present inventors thus tested whether the Gwt1 protein was involved in this transacylation reaction using the method described below.

[0035] The membrane fraction preparation (300 µg protein) was diluted with a buffer consisting of 50 mM Tris-HCl (pH7.5), 2 mM MgCl₂, 2 mM MnCl₂, 1 mM ATP, 1 mM Coenzyme A, 21 µg/ml tunicamycin, 10 µM nikkomycin Z, and 0.5 mM Dithiothreitol. The solution was adjusted to a total of 140 µl for use as a reaction solution. After incubating the solution at 24°C for 15 minutes, 15 µCi UDP-[¹⁴C]GlcNAc was added to the tube and then incubated at 24°C for another one hour. 1 ml of chloroform:methanol (1:2) was added to the solution and stirred to stop the reaction. Then, lipid was extracted from the solution, dried, and desalted by butanol extraction. Acylated GPI (GlcN-(acyl)PI), non-acylated GPI (GlcN-PI), and GPI which was neither acylated nor deacetylated (GlcNAc-PI) were separated by thin-layer chromatography (HCl₃/CH₃OH/1 M NH₄OH (10:10:3)). Each spot was detected by autoradiography.

[0036] As a result, as shown in Fig. 2, a spot for acylated GPI was not detected in the GWT1 gene-deficient strain (Δ gwt1), while it was detected in the wild-type strain. The spot for acylated GPI was also detected in the GWT1 gene-introduced Δ gwt1 strain, showing that this strain had recovered ability to acylate. These findings indicate that the Gwt1 protein is an enzyme that catalyzes transacylation to GPI.

[0037] The above-described results suggest that the intensity of the spot for acylated GlcN-(acyl)PI is reduced or disappears when a compound having the activity of inhibiting the activity of GWT1 gene products is present in a system for assaying GPI synthase activity. Accordingly, compounds inhibiting the enzymatic activity of a GWT1 gene product, as well as compounds inhibiting the synthesis of fungal cell walls, can be screened using the intensity of GlcN-(acyl)PI spots as an indicator.

[0038] [Effects of the Invention]

The present invention makes it possible to screen for compounds that inhibit the transport of GPI-anchored proteins to fungal cell walls by using a simple assay of transacylation activity.

5 [0039] [Sequence Listing]

<110> Eisai Co., Ltd.

National Institute of Advanced Industrial Science and Technology

10 <120> Method for a screening of an inhibitor of GWT1 gene product

<130>

<160> 18

15 <170> PatentIn Ver. 2.0

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5

10

15

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20

25

30

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35

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40

45

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	Asn	Pro	Ile	Tyr	Asn	Lys	Lys	Lys	Met	Ile	Thr	Gln	Arg	Phe	Gln	Leu	
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	tct tca ccc cat aga caa aat gat aca aaa gaa gat aaa tcg gac gaa	336
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	Ala Phe Ser Leu Phe Ile Ser Asn Leu Ser Phe Leu Gln Pro Ile Ser	
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	gct acg ttt tta tta tgt tat gac tta att gaa aaa ttt atc ccg ggg	1248
	Ala Thr Phe Leu Leu Cys Tyr Asp Leu Ile Glu Lys Phe Ile Pro Gly	
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 1 5 10 15
 act ggt ggc aca att gaa gaa att tat gct gta acc agt ata gca tta 96
 Thr Gly Gly Thr Ile Glu Glu Ile Tyr Ala Val Thr Ser Ile Ala Leu

25 20 25 30
 tca tct tat ttg tcc ttt aga ttg ttg aaa aag tct ctt ggt gat tta 144
 Ser Ser Tyr Leu Ser Phe Arg Leu Leu Lys Lys Ser Leu Gly Asp Leu
 35 40 45
 gct ttg att tac gac tac att ctt aat gtg ttg aca att cta gca tcc 192

30 Ala Leu Ile Tyr Asp Tyr Ile Leu Asn Val Leu Thr Ile Leu Ala Ser
 50 55 60
 att act gtt tat agc aac agc cct tct tat ttg cat tat ttt att gtt 240
 Ile Thr Val Tyr Ser Asn Ser Pro Ser Tyr Leu His Tyr Phe Ile Val
 65 70 75 80

35 att cca tca tta gtt ata tat cta gtg aat tac cat gtt gag aaa cca 288
 Ile Pro Ser Leu Val Ile Tyr Leu Val Asn Tyr His Val Glu Lys Pro

		85		90		95		
	tct tca ccc cat aga caa aat gat aca aaa gaa gat aaa tcg gac gaa	336						
	Ser Ser Pro His Arg Gln Asn Asp Thr Lys Glu Asp Lys Ser Asp Glu							
	100		105		110			
5	cta ttg ccg aga aaa caa ttt ata aca gcc tat cgt tct caa atg ttg	384						
	Leu Leu Pro Arg Lys Gln Phe Ile Thr Ala Tyr Arg Ser Gln Met Leu							
	115		120		125			
	ata att act aat cta gct ata tta gct gtt gat ttt cct att ttc cca	432						
	Ile Ile Thr Asn Leu Ala Ile Leu Ala Val Asp Phe Pro Ile Phe Pro							
10	130		135		140			
	aga aga ttt gcc aaa gtg gaa aca tgg ggc acg tca atg atg gat tta	480						
	Arg Arg Phe Ala Lys Val Glu Thr Trp Gly Thr Ser Met Met Asp Leu							
	145		150		155		160	
	gga gtt ggg tcg ttt gtg ttc tcc atg ggg ttg gct aat tct cga caa	528						
15	Gly Val Gly Ser Phe Val Phe Ser Met Gly Leu Ala Asn Ser Arg Gln							
	165		170		175			
	ttg atc aag aac cac acc gac aat tac aaa ttt agt tgg aag agt tat	576						
	Leu Ile Lys Asn His Thr Asp Asn Tyr Lys Phe Ser Trp Lys Ser Tyr							
	180		185		190			
20	ttg aaa aca atc aag cag aac ttt atc aag tca gtg cct ata ctt gtt	624						
	Leu Lys Thr Ile Lys Gln Asn Phe Ile Lys Ser Val Pro Ile Leu Val							
	195		200		205			
	tta gga gct att cgt ttt gtt agt gtt aag caa ttg gac tat cag gaa	672						
	Leu Gly Ala Ile Arg Phe Val Ser Val Lys Gln Leu Asp Tyr Gln Glu							
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	cac gaa aca gag tat gga atc cat tgg aat ttt ttc ttc aca tta ggg	720						
	His Glu Thr Glu Tyr Gly Ile His Trp Asn Phe Phe Phe Thr Leu Gly							
	225		230		235		240	
	ttc ttg cca att gta ttg gga ata tta gac ccg gtg ttg aat ttg gtt	768						
30	Phe Leu Pro Ile Val Leu Gly Ile Leu Asp Pro Val Leu Asn Leu Val							
	245		250		255			
	cca cgc ttc ata ata gga att ggt atc tca att ggt tat gag gta gcg	816						
	Pro Arg Phe Ile Ile Gly Ile Gly Ile Ser Ile Gly Tyr Glu Val Ala							
	260		265		270			
35	ttg aat aag act ggt ttg ttg aag ttc att ttg agc agc gaa aac aga	864						
	Leu Asn Lys Thr Gly Leu Leu Lys Phe Ile Leu Ser Ser Glu Asn Arg							

	275	280	285	
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	Leu Glu Ser Leu Ile Ala Met Asn Lys Glu Gly Ile Phe Ser Phe Ile			
	290	295	300	
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	Gly Tyr Leu Cys Ile Phe Ile Ile Gly Gln Ser Phe Gly Ser Phe Val			
	305	310	315	320
	tta aca ggc tac aaa aca aag aac aac tta ata acc att agc aaa att	1008		
	Leu Thr Gly Tyr Lys Thr Lys Asn Asn Leu Ile Thr Ile Ser Lys Ile			
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	cgt att tca aaa aaa caa cac aag aaa gag ctg ctg ctg ttt ttc tca	1056		
	Arg Ile Ser Lys Lys Gln His Lys Lys Glu Leu Leu Leu Phe Phe Ser			
	340	345	350	
	gtc gcc act act cag gga tta tat ttg gca tgt atc ttc tat cac tta	1104		
15	Val Ala Thr Thr Gln Gly Leu Tyr Leu Ala Cys Ile Phe Tyr His Leu			
	355	360	365	
	gct ttc agt ttg ttc atc agc aac tta tca ttc ttg caa cca att tca	1152		
	Ala Phe Ser Leu Phe Ile Ser Asn Leu Ser Phe Leu Gln Pro Ile Ser			
	370	375	380	
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	Arg Arg Leu Ala Asn Phe Pro Tyr Val Met Trp Val Val Ser Tyr Asn			
	385	390	395	400
	gct acg ttt tta tta tgt tat gac tta att gaa aaa ttt atc ccg ggg	1248		
	Ala Thr Phe Leu Leu Cys Tyr Asp Leu Ile Glu Lys Phe Ile Pro Gly			
25	405	410	415	
	aac ctt act tct act gta ttg gac tct att aat aac aat ggt tta ttt	1296		
	Asn Leu Thr Ser Thr Val Leu Asp Ser Ile Asn Asn Asn Gly Leu Phe			
	420	425	430	
	atc ttc ttg gtc agc aat tta tta aca ggg ttt att aac atg tcc atc	1344		
30	Ile Phe Leu Val Ser Asn Leu Leu Thr Gly Phe Ile Asn Met Ser Ile			
	435	440	445	
	aac act ttg gaa act agc aat aaa atg gca gtg att atc ttg att ggc	1392		
	Asn Thr Leu Glu Thr Ser Asn Lys Met Ala Val Ile Ile Leu Ile Gly			
	450	455	460	
35	tat agt ctt act tgg aca ttg ctc gcc tta tat ttg gat aag agg aag	1440		
	Tyr Ser Leu Thr Trp Thr Leu Leu Ala Leu Tyr Leu Asp Lys Arg Lys			

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 atc tac atc aag ctt tag 1458
 Ile Tyr Ile Lys Leu
 485
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 35 40 45
 Ala Leu Ile Tyr Asp Tyr Ile Leu Asn Val Leu Thr Ile Leu Ala Ser
 20 50 55 60
 Ile Thr Val Tyr Ser Asn Ser Pro Ser Tyr Leu His Tyr Phe Ile Val
 65 70 75 80
 Ile Pro Ser Leu Val Ile Tyr Leu Val Asn Tyr His Val Glu Lys Pro
 85 90 95
 25 Ser Ser Pro His Arg Gln Asn Asp Thr Lys Glu Asp Lys Ser Asp Glu
 100 105 110
 Leu Leu Pro Arg Lys Gln Phe Ile Thr Ala Tyr Arg Ser Gln Met Leu
 115 120 125
 Ile Ile Thr Asn Leu Ala Ile Leu Ala Val Asp Phe Pro Ile Phe Pro
 30 130 135 140
 Arg Arg Phe Ala Lys Val Glu Thr Trp Gly Thr Ser Met Met Asp Leu
 145 150 155 160
 Gly Val Gly Ser Phe Val Phe Ser Met Gly Leu Ala Asn Ser Arg Gln
 165 170 175
 35 Leu Ile Lys Asn His Thr Asp Asn Tyr Lys Phe Ser Trp Lys Ser Tyr
 180 185 190

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	225				230				235								240	
	Phe	Leu	Pro	Ile	Val	Leu	Gly	Ile	Leu	Asp	Pro	Val	Leu	Asn	Leu	Val		
	245				250				255									
	Pro	Arg	Phe	Ile	Ile	Gly	Ile	Gly	Ile	Ser	Ile	Gly	Tyr	Glu	Val	Ala		
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	Leu	Asn	Lys	Thr	Gly	Leu	Leu	Lys	Phe	Ile	Leu	Ser	Ser	Glu	Asn	Arg		
	275				280				285									
	Leu	Glu	Ser	Leu	Ile	Ala	Met	Asn	Lys	Glu	Gly	Ile	Phe	Ser	Phe	Ile		
	290				295				300									
15	Gly	Tyr	Leu	Cys	Ile	Phe	Ile	Ile	Gly	Gln	Ser	Phe	Gly	Ser	Phe	Val		
	305				310				315								320	
	Leu	Thr	Gly	Tyr	Lys	Thr	Lys	Asn	Asn	Leu	Ile	Thr	Ile	Ser	Lys	Ile		
	325				330				335									
	Arg	Ile	Ser	Lys	Lys	Gln	His	Lys	Lys	Glu	Leu	Leu	Leu	Phe	Phe	Ser		
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	355				360				365									
	Ala	Phe	Ser	Leu	Phe	Ile	Ser	Asn	Leu	Ser	Phe	Leu	Gln	Pro	Ile	Ser		
	370				375				380									
25	Arg	Arg	Leu	Ala	Asn	Phe	Pro	Tyr	Val	Met	Trp	Val	Val	Ser	Tyr	Asn		
	385				390				395								400	
	Ala	Thr	Phe	Leu	Leu	Cys	Tyr	Asp	Leu	Ile	Glu	Lys	Phe	Ile	Pro	Gly		
	405				410				415									
	Asn	Leu	Thr	Ser	Thr	Val	Leu	Asp	Ser	Ile	Asn	Asn	Asn	Gly	Leu	Phe		
30	420				425				430									
	Ile	Phe	Leu	Val	Ser	Asn	Leu	Leu	Thr	Gly	Phe	Ile	Asn	Met	Ser	Ile		
	435				440				445									
	Asn	Thr	Leu	Glu	Thr	Ser	Asn	Lys	Met	Ala	Val	Ile	Ile	Leu	Ile	Gly		
	450				455				460									
35	Tyr	Ser	Leu	Thr	Trp	Thr	Leu	Leu	Ala	Leu	Tyr	Leu	Asp	Lys	Arg	Lys		
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Ile Tyr Ile Lys Leu
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5 <211> 1380

<212> DNA

<213> Schizosaccharomyces pombe

<220>

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	Ser Ser Ser Ile Glu Thr Cys Gly Leu Leu Leu Ile Gly Ile Ala Cys	
	20 25 30	
20	aac gtt ttg tgg gta aac atg act gcg aga aac atc tta ccc aaa ggg	144
	Asn Val Leu Trp Val Asn Met Thr Ala Arg Asn Ile Leu Pro Lys Gly	
	35 40 45	
	aat ctt ggg ttt ctt gtt gag ttt ttc atc ttt tgc tta att cca tta	192
	Asn Leu Gly Phe Leu Val Glu Phe Phe Ile Phe Cys Leu Ile Pro Leu	
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	Phe Val Ile Tyr Val Ser Ser Lys Val Gly Val Phe Thr Leu Cys Ile	
	65 70 75 80	
	gcc tct ttt ttg cct tcc ttc gtc ctt cat gtt ata agt cca att aat	288
30	Ala Ser Phe Leu Pro Ser Phe Val Leu His Val Ile Ser Pro Ile Asn	
	85 90 95	
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	Trp Asp Val Leu Arg Arg Lys Pro Gly Cys Cys Leu Thr Lys Lys Asn	
	100 105 110	
35	gaa aat act ttt gat cga cga att gct gga gtc aca ttt tat cgt tct	384
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	115	120	125	
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	130	135	140	
5	ctt ttc ccg agg aga tat gcc aaa gtt gaa acc tgg gga aca tca ctg	480		
	Leu Phe Pro Arg Arg Tyr Ala Lys Val Glu Thr Trp Gly Thr Ser Leu			
	145	150	155	160
	atg gat ctt ggt gtt gga tct ttc atg ttt tct tca ggt act gtg gct	528		
	Met Asp Leu Gly Val Gly Ser Phe Met Phe Ser Ser Gly Thr Val Ala			
10	165	170	175	
	gga cgg aaa aat gac att aaa aaa cca aat gcg ttt aaa aat gta ttg	576		
	Gly Arg Lys Asn Asp Ile Lys Lys Pro Asn Ala Phe Lys Asn Val Leu			
	180	185	190	
	tgg aat tct ttc atc ctt ttg att tta gga ttt gcg cgc atg ttt tta	624		
15	Trp Asn Ser Phe Ile Leu Leu Ile Leu Gly Phe Ala Arg Met Phe Leu			
	195	200	205	
	acg aaa agc atc aat tac caa gaa cat gta agc gaa tat ggc atg cat	672		
	Thr Lys Ser Ile Asn Tyr Gln Glu His Val Ser Glu Tyr Gly Met His			
	210	215	220	
20	tgg aac ttt ttt ttc acc cta ggt ttc atg gct ctt ggc gta ttt ttt	720		
	Trp Asn Phe Phe Phe Thr Leu Gly Phe Met Ala Leu Gly Val Phe Phe			
	225	230	235	240
	ttt cgt cgt tct tta aaa aaa gtc tcc tat ttt aat tta gca acc ttc	768		
	Phe Arg Arg Ser Leu Lys Lys Val Ser Tyr Phe Asn Leu Ala Thr Phe			
25	245	250	255	
	att act ctt ctt cat cat tgt ttg ctt gtt tta acc cct ttc caa aaa	816		
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	260	265	270	
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	275	280	285	
	ggt att gct tct ctt ccc gga tac att gct att tac ttt tat gga atg	912		
	Gly Ile Ala Ser Leu Pro Gly Tyr Ile Ala Ile Tyr Phe Tyr Gly Met			
	290	295	300	
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	Tyr Thr Gly Ser Val Val Leu Ala Asp Arg Pro Leu Met Tyr Thr Arg			

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	Ala	Glu	Ser	Trp	Lys	Arg	Phe	Gln	Arg	Leu	Leu	Phe	Pro	Leu	Cys	Ile	
					325					330					335		
5	ttg	tta	gtg	ttg	tat	ctt	gtg	tct	aac	ttt	ttg	tca	gtt	ggg	ggt	tct	1056
	Leu	Leu	Val	Leu	Tyr	Leu	Val	Ser	Asn	Phe	Leu	Ser	Val	Gly	Val	Ser	
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	cgc	cga	ctt	gct	aat	acg	cct	tat	gtt	gcg	aat	gtt	gcc	ttt	atc	aat	1104
	Arg	Arg	Leu	Ala	Asn	Thr	Pro	Tyr	Val	Ala	Asn	Val	Ala	Phe	Ile	Asn	
10					355					360					365		
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	Met	Phe	Phe	Leu	Thr	Ile	Tyr	Ile	Leu	Ile	Asp	Ala	Tyr	Leu	Phe	Pro	
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<211> 459

<212> PRT

<213> Schizosaccharomyces pombe

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				20					25					30		
5	Asn	Val	Leu	Trp	Val	Asn	Met	Thr	Ala	Arg	Asn	Ile	Leu	Pro	Lys	Gly
			35					40					45			
	Asn	Leu	Gly	Phe	Leu	Val	Glu	Phe	Phe	Ile	Phe	Cys	Leu	Ile	Pro	Leu
		50					55					60				
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10	65					70					75				80	
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				85					90					95		
	Trp	Asp	Val	Leu	Arg	Arg	Lys	Pro	Gly	Cys	Cys	Leu	Thr	Lys	Lys	Asn
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		130					135				140					
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			165						170				175			
	Gly	Arg	Lys	Asn	Asp	Ile	Lys	Lys	Pro	Asn	Ala	Phe	Lys	Asn	Val	Leu
			180						185				190			
25	Trp	Asn	Ser	Phe	Ile	Leu	Leu	Ile	Leu	Gly	Phe	Ala	Arg	Met	Phe	Leu
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		210					215					220				
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	Phe	Arg	Arg	Ser	Leu	Lys	Lys	Val	Ser	Tyr	Phe	Asn	Leu	Ala	Thr	Phe
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			260					265				270				
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		275						280				285				

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 Tyr Thr Gly Ser Val Val Leu Ala Asp Arg Pro Leu Met Tyr Thr Arg
 305 310 315 320
 5 Ala Glu Ser Trp Lys Arg Phe Gln Arg Leu Leu Phe Pro Leu Cys Ile
 325 330 335
 Leu Leu Val Leu Tyr Leu Val Ser Asn Phe Leu Ser Val Gly Val Ser
 340 345 350
 Arg Arg Leu Ala Asn Thr Pro Tyr Val Ala Asn Val Ala Phe Ile Asn
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 370 375 380
 Ser Ser Val Pro Tyr Gly Ser Arg Val Pro Lys Leu Leu Glu Asp Ala
 385 390 395 400
 15 Asn Asn Asn Gly Leu Leu Val Phe Leu Ile Ala Asn Val Leu Thr Gly
 405 410 415
 Val Val Asn Leu Ser Phe Asp Thr Leu His Ser Ser Asn Ala Lys Gly
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5	att cct ctg ctc gtc ctc ggc ctg att cgg cta tac agc gtc aaa ggc	726						
	Ile Pro Leu Leu Val Leu Gly Leu Ile Arg Leu Tyr Ser Val Lys Gly							
		220		225		230		
	ttg gac tat gcg gag cac gtc acc gag tac ggc gta cat tgg aac ttc	774						
	Leu Asp Tyr Ala Glu His Val Thr Glu Tyr Gly Val His Trp Asn Phe							
10		235		240		245		
	ttc ttt aca ttg ggt ctt ttg cct ccg ttc gtg gag gtc ttc gac gcc	822						
	Phe Phe Thr Leu Gly Leu Leu Pro Pro Phe Val Glu Val Phe Asp Ala							
		250		255		260		
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	265		270		275		280	
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	Val Leu Tyr Gln Val Ala Leu Glu Ser Thr Asp Leu Lys Ser Tyr Ile							
		285		290		295		
20	ctc gtc tcc cct cgt ggg cca agc tta ctg tcc aag aat cgt gaa ggc	966						
	Leu Val Ser Pro Arg Gly Pro Ser Leu Leu Ser Lys Asn Arg Glu Gly							
		300		305		310		
	gtc ttc tcc ttc tca ggt tat ctc gcg att ttt ctt gct ggt cgt gcg	1014						
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25		315		320		325		
	atc ggc att cgg ata atc cct cgc gga act tct ttc tca aga agc cca	1062						
	Ile Gly Ile Arg Ile Ile Pro Arg Gly Thr Ser Phe Ser Arg Ser Pro							
		330		335		340		
	gaa cag gcc agg aga cgg gtc ctg atc agc ctt ggc gtg caa gcg tta	1110						
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	345		350		355		360	
	gtg tgg acc act ctt ttt gtg ttg aac tcc act tat gcg atg gga tac	1158						
	Val Trp Thr Thr Leu Phe Val Leu Asn Ser Thr Tyr Ala Met Gly Tyr							
		365		370		375		
35	gga gct aat atc cct gtc tcc cgc cgc ctc gct aac atg ccc tat gtc	1206						
	Gly Ala Asn Ile Pro Val Ser Arg Arg Leu Ala Asn Met Pro Tyr Val							

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	Leu Trp Val Ser Ala Phe Asn Thr Ala Gln Leu Phe Val Phe Cys Leu			
	395	400	405	
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	gga gct gtg aat ctg agc atc tcc aca att gat gct aat aca gcg cag	1446		
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Glu Ala Phe Val Ser Gly Leu Ala Gly Gly Ser Ile Leu Glu Ile Asn
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20 cgcctacatt gtttctcgac taaccgagtc tctttgcgat caatcag gta tcc gtt      207
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25 Phe Leu Trp Ser Ile Leu Gln Ser Arg Leu Ser Phe Phe Thr Pro Tyr
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	Leu	Ala	Gly	Arg	Ala	Ile	Gly	Ile	Arg	Ile	Ile	Pro	Arg	Gly	Thr	Ser		
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 Ala Thr Tyr Ala
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	Thr	Ser	Pro	Leu	Val	Phe	Thr	Ser	Phe	Leu	Ser	Ile	Ile	Ser	Leu	Ala	
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	His Val Thr Glu Tyr Gly Val His Trp Asn Phe Phe Phe Thr Leu Ala			
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	Leu Val Pro Val Leu Ala Val Gly Ile Arg Pro Leu Thr Gln Trp Leu			
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	cgc tgg agt gtg ctt ggg gta atc atc tct ttg ctg cat cag ctg tgg	1317		
30	Arg Trp Ser Val Leu Gly Val Ile Ile Ser Leu Leu His Gln Leu Trp			
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	Ser Leu Pro Pro Arg Arg Glu Arg Val Val Ser Glu Thr Asn Glu Glu			
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	Thr Leu Thr Ile Ser Cys Val Gly Trp Ile Leu Lys Gly Arg Arg Ile			

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 Thr Tyr Ala Leu Trp Ile Ala Leu Ser Pro Tyr Ile Arg His Gly Leu
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 Leu Asn Asn Tyr Leu Ile Cys Val Leu Pro Leu Leu Phe Gly Val Thr
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	Leu Thr Val Tyr Arg Ala His Met Met Leu Met Thr Val Ile Cys Ile	
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	Asp Phe Gly Thr Ser Leu Met Asp Val Gly Val Gly Ser Phe Val Phe	
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	Ser Leu Gly Leu Val Ser Thr Lys Ser Leu Ser Pro Pro Pro Thr	
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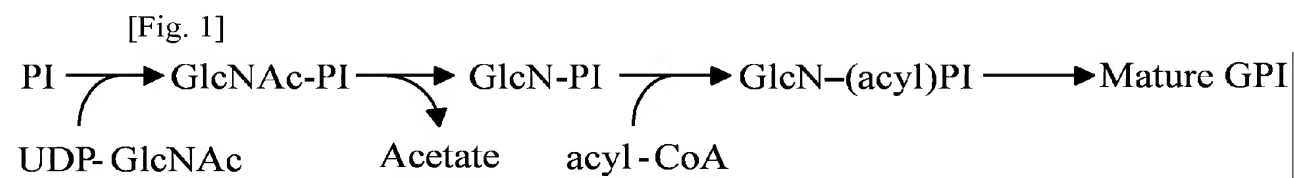
[Brief Description of the Drawings]

[Fig. 1] The GPI biosynthesis pathway is shown.

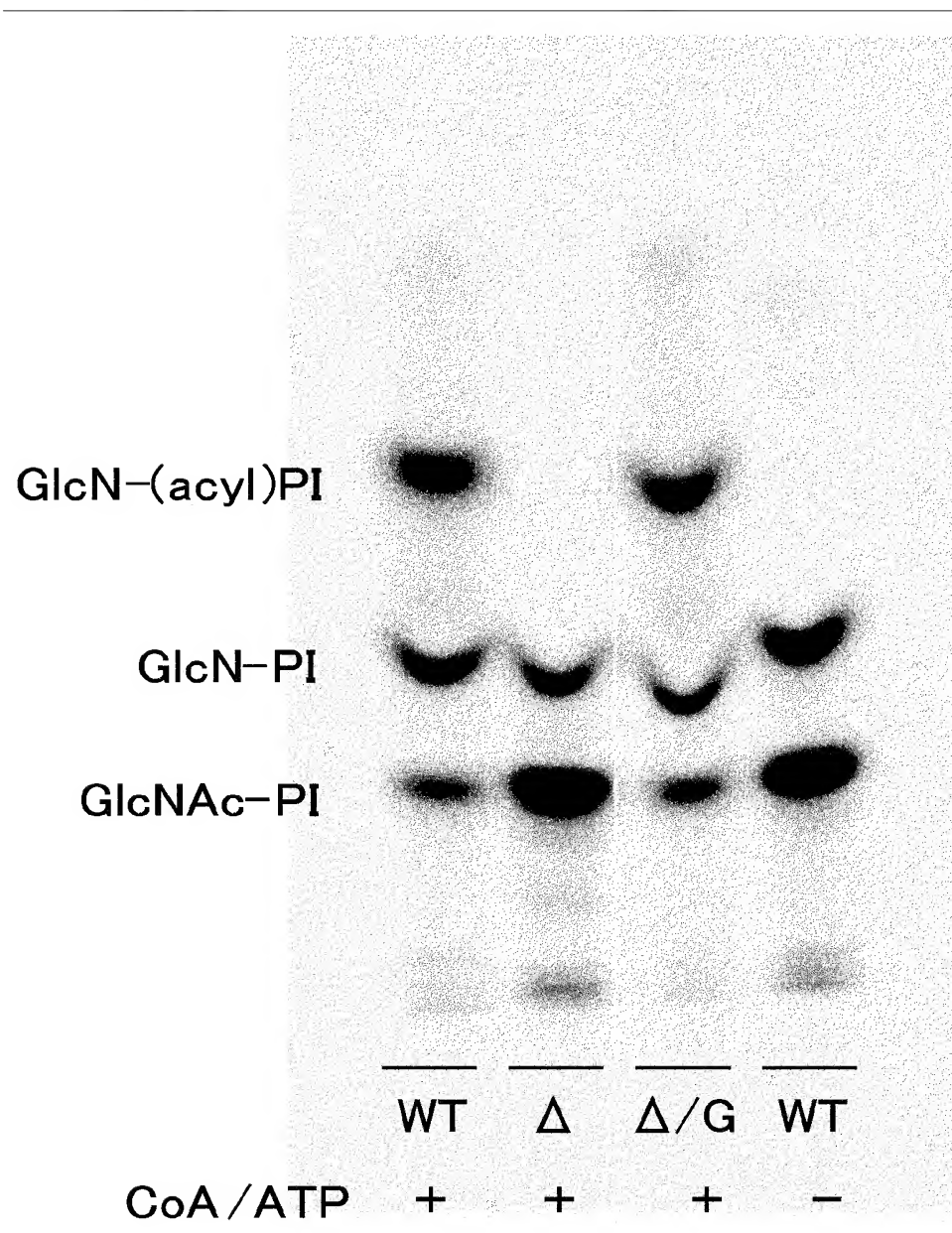
[Fig. 2] A photograph showing the inhibition of binding of labeled CompoundB2 to the membrane fraction by the subject compound is depicted.

5

[Document Name] Drawings



[Fig. 2]



[Document Name] Abstract

[Abstract]

[Problems to be Solved]

5 An objective is to develop antifungal agents for preventing pathogenic fungi from
exerting pathogenicity, by inhibiting the synthesis of fungal cell walls, as well as by inhibiting
fungal cell adhesion to host cells, by inhibition of the transport of GPI-anchored proteins to
fungal cell walls.

[Means to Solve the Problems]

10 The present invention enables screening for compounds that inhibit the transport of
GPI-anchored proteins to fungal cell walls, using a simple binding assay using membrane
fraction expressing GWT1 protein. New antifungal agents can be created that inhibit the
synthesis of fungal cell walls and also inhibit adhesion to host cells by inhibiting the transport of
GPI-anchored proteins to fungal cell walls.

[Selected Drawings] None

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